

# p-Y-STAT1,3,5 dimerize

May, B., Touw, IP.

European Bioinformatics Institute, New York University Langone Medical Center, Ontario Institute for Cancer Research, Oregon Health and Science University.

The contents of this document may be freely copied and distributed in any media, provided the authors, plus the institutions, are credited, as stated under the terms of <u>Creative Commons Attribution 4.0 International (CC BY 4.0)</u> <u>License</u>. For more information see our <u>license</u>.

04/05/2024

# Introduction

Reactome is open-source, open access, manually curated and peer-reviewed pathway database. Pathway annotations are authored by expert biologists, in collaboration with Reactome editorial staff and cross-referenced to many bioinformatics databases. A system of evidence tracking ensures that all assertions are backed up by the primary literature. Reactome is used by clinicians, geneticists, genomics researchers, and molecular biologists to interpret the results of high-throughput experimental studies, by bioinformaticians seeking to develop novel algorithms for mining knowledge from genomic studies, and by systems biologists building predictive models of normal and disease variant pathways.

The development of Reactome is supported by grants from the US National Institutes of Health (P41 HG003751), University of Toronto (CFREF Medicine by Design), European Union (EU STRP, EMI-CD), and the European Molecular Biology Laboratory (EBI Industry program).

### Literature references

- Fabregat, A., Sidiropoulos, K., Viteri, G., Forner, O., Marin-Garcia, P., Arnau, V. et al. (2017). Reactome pathway analysis: a high-performance in-memory approach. *BMC bioinformatics, 18,* 142. 7
- Sidiropoulos, K., Viteri, G., Sevilla, C., Jupe, S., Webber, M., Orlic-Milacic, M. et al. (2017). Reactome enhanced pathway visualization. *Bioinformatics*, 33, 3461-3467. A
- Fabregat, A., Jupe, S., Matthews, L., Sidiropoulos, K., Gillespie, M., Garapati, P. et al. (2018). The Reactome Pathway Knowledgebase. *Nucleic Acids Res, 46*, D649-D655. ↗
- Fabregat, A., Korninger, F., Viteri, G., Sidiropoulos, K., Marin-Garcia, P., Ping, P. et al. (2018). Reactome graph database: Efficient access to complex pathway data. *PLoS computational biology*, *14*, e1005968. *オ*

This document contains 1 reaction (see Table of Contents)

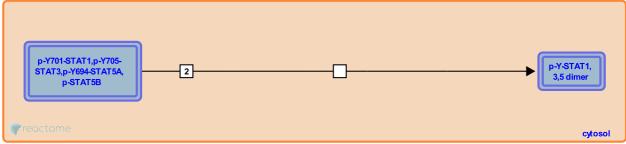
## p-Y-STAT1,3,5 dimerize 🛪

Stable identifier: R-HSA-9674542

Type: transition

#### Compartments: cytosol

Inferred from: p-Y-Stat1,3,5 dimerize (Mus musculus)



After being phosphorylated, STAT1, STAT3, and STAT5 (STAT5A and STAT5B) form homodimers (Tian et al. 1994, Tian et al. 1996, Ward et al. 1999, also inferred from mouse homologs). Phospho-STAT1 and phospho-STAT3 can also form heterodimers (Tian et al. 1994, Ward et al. 1999, also inferred from mouse homologs).

#### Literature references

- Tian, SS., Seidel, HM., Rosen, J., Stein, RB., Lamb, P. (1994). Rapid activation of the STAT3 transcription factor by granulocyte colony-stimulating factor. *Blood*, *84*, 1760-4.
- Schelen, AM., Smith, L., Ward, AC., Touw, IP., Hermans, MH., Antonissen, C. et al. (1999). Tyrosine-dependent and independent mechanisms of STAT3 activation by the human granulocyte colony-stimulating factor (G-CSF) receptor are differentially utilized depending on G-CSF concentration. *Blood*, *93*, 113-24.
- Tian, SS., Rosen, J., Tapley, P., Stein, RB., Sincich, C., Lamb, P. (1996). Multiple signaling pathways induced by granulocyte colony-stimulating factor involving activation of JAKs, STAT5, and/or STAT3 are required for regulation of three distinct classes of immediate early genes. *Blood, 88*, 4435-44.

#### **Editions**

2020-01-13	Authored, Edited	May, B.
2020-12-12	Reviewed	Touw, IP.