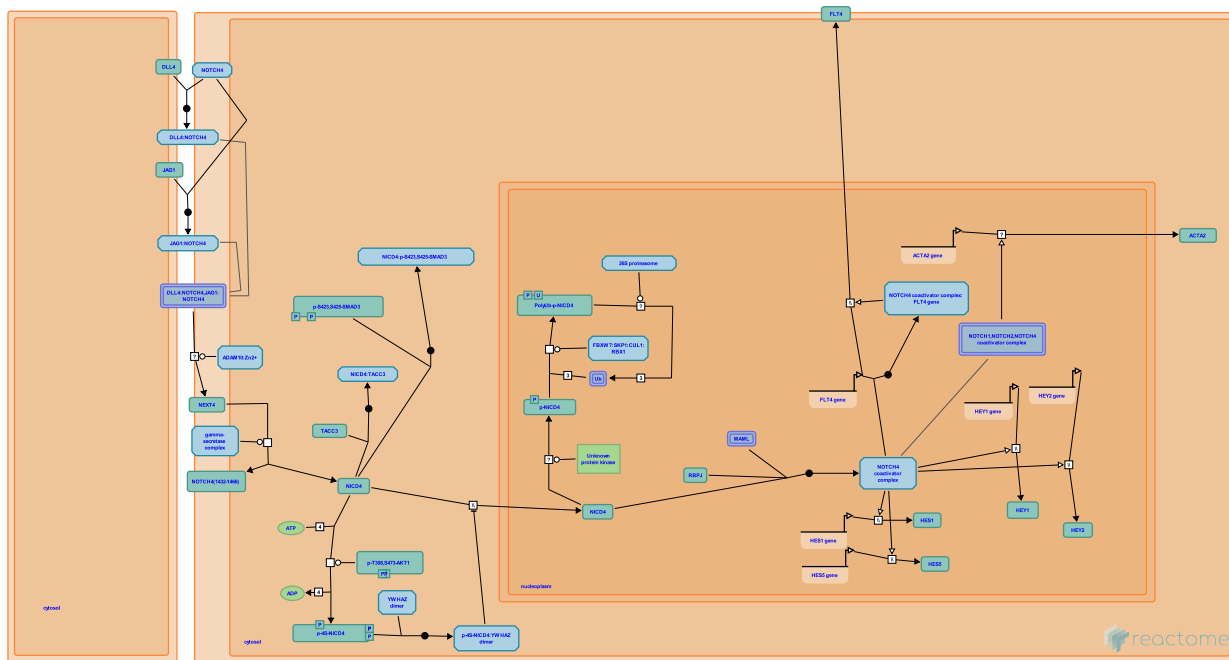


Signaling by NOTCH4



Haw, R., Jassal, B., Joutel, A., Orlic-Milacic, M.

European Bioinformatics Institute, New York University Langone Medical Center, Ontario Institute for Cancer Research, Oregon Health and Science University.

The contents of this document may be freely copied and distributed in any media, provided the authors, plus the institutions, are credited, as stated under the terms of [Creative Commons Attribution 4.0 International \(CC BY 4.0\) License](https://creativecommons.org/licenses/by/4.0/). For more information see our [license](https://reactome.org/page/faq).

This is just an excerpt of a full-length report for this pathway. To access the complete report, please download it at the [Reactome Textbook](https://reactome.org/page/faq).

10/04/2024

Introduction

Reactome is open-source, open access, manually curated and peer-reviewed pathway database. Pathway annotations are authored by expert biologists, in collaboration with Reactome editorial staff and cross-referenced to many bioinformatics databases. A system of evidence tracking ensures that all assertions are backed up by the primary literature. Reactome is used by clinicians, geneticists, genomics researchers, and molecular biologists to interpret the results of high-throughput experimental studies, by bioinformaticians seeking to develop novel algorithms for mining knowledge from genomic studies, and by systems biologists building predictive models of normal and disease variant pathways.

The development of Reactome is supported by grants from the US National Institutes of Health (P41 HG003751), University of Toronto (CFREF Medicine by Design), European Union (EU STRP, EMI-CD), and the European Molecular Biology Laboratory (EBI Industry program).

Literature references

Fabregat, A., Sidiropoulos, K., Viteri, G., Forner, O., Marin-Garcia, P., Arnau, V. et al. (2017). Reactome pathway analysis: a high-performance in-memory approach. *BMC bioinformatics*, 18, 142. [↗](#)

Sidiropoulos, K., Viteri, G., Sevilla, C., Jupe, S., Webber, M., Orlic-Milacic, M. et al. (2017). Reactome enhanced pathway visualization. *Bioinformatics*, 33, 3461-3467. [↗](#)

Fabregat, A., Jupe, S., Matthews, L., Sidiropoulos, K., Gillespie, M., Garapati, P. et al. (2018). The Reactome Pathway Knowledgebase. *Nucleic Acids Res*, 46, D649-D655. [↗](#)

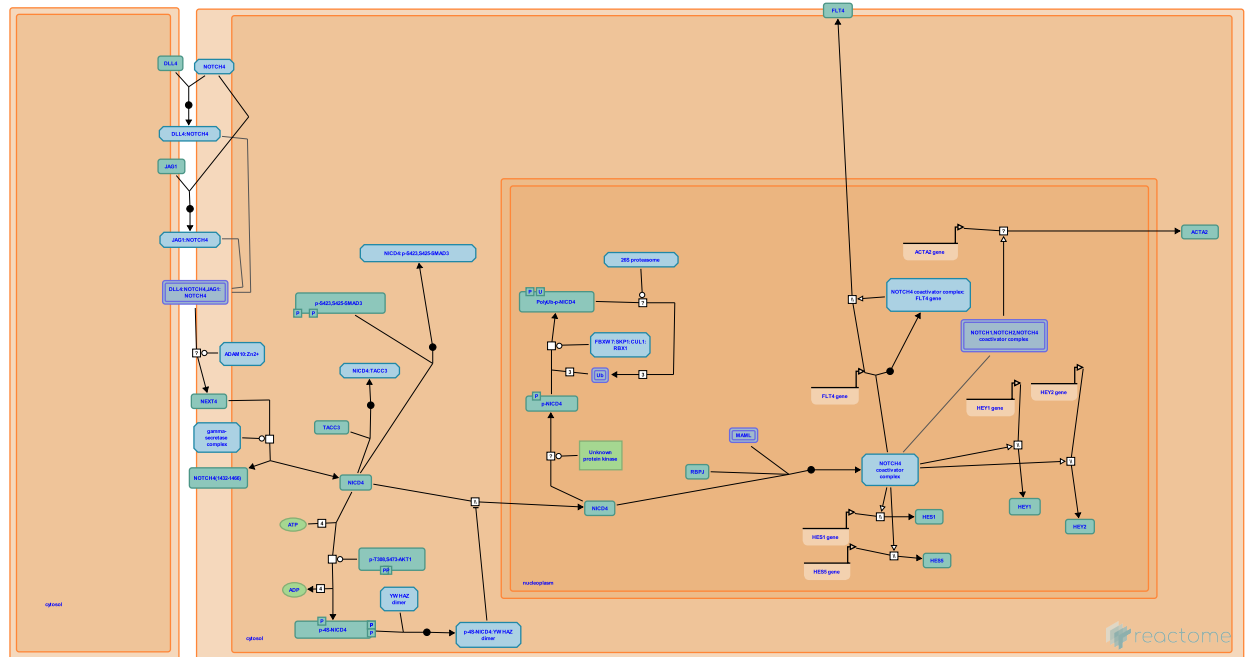
Fabregat, A., Korninger, F., Viteri, G., Sidiropoulos, K., Marin-Garcia, P., Ping, P. et al. (2018). Reactome graph database: Efficient access to complex pathway data. *PLoS computational biology*, 14, e1005968. [↗](#)

Reactome database release: 88

This document contains 4 pathways ([see Table of Contents](#))

Signaling by NOTCH4 ↗

Stable identifier: R-HSA-9013694



The NOTCH4 gene locus was discovered as a frequent site of insertion for the proviral genome of the mouse mammary tumor virus (MMTV) (Gallahan and Callahan 1987). MMTV-insertion results in aberrant expression of the mouse mammary tumor gene int-3, which was subsequently discovered to represent the intracellular domain of Notch4 (Robbins et al. 1992, Uyttendaele et al. 1996).

NOTCH4 is prevalently expressed in endothelial cells (Uyttendaele et al. 1996). DLL4 and JAG1 act as ligands for NOTCH4 in human endothelial cells (Shawber et al. 2003, Shawber et al. 2007), but DLL4- and JAG1-mediated activation of NOTCH4 have not been confirmed in all cell types tested (Aste-Amezaga et al. 2010, James et al. 2014). The gamma secretase complex cleaves activated NOTCH4 receptor to release the intracellular domain fragment (NICD4) (Saxena et al. 2001, Das et al. 2004). NICD4 traffics to the nucleus where it acts as a transcription factor and stimulates expression of NOTCH target genes HES1, HES5, HEY1 and HEY2, as well as VEGFR3 and ACTA2 (Lin et al. 2002, Raafat et al. 2004, Tsunematsu et al. 2004, Shawber et al. 2007, Tang et al. 2008, Bargo et al. 2010). NOTCH4 signaling can be downregulated by AKT1 phosphorylation-induced cytoplasmic retention (Ramakrishnan et al. 2015) as well as proteasome-dependent degradation upon FBXW7-mediated ubiquitination (Wu et al. 2001, Tsunematsu et al. 2004).

NOTCH4 was reported to inhibit NOTCH1 signaling in-cis, by binding to NOTCH1 and interfering with the S1 cleavage of NOTCH1, thus preventing production of functional NOTCH1 heterodimers at the cell surface (James et al. 2014).

NOTCH4 is involved in development of the vascular system. Overexpression of constitutively active Notch4 in mouse embryonic vasculature results in abnormal vessel structure and patterning (Uyttendaele et al. 2001). NOTCH4 may act to inhibit apoptosis of endothelial cells (MacKenzie et al. 2004).

Expression of int-3 interferes with normal mammary gland development in mice and promotes tumorigenesis. The phenotype of mice expressing int-3 in mammary glands is dependent on the presence of Rbpj (Raafat et al. 2009). JAG1 and NOTCH4 are upregulated in human ER+ breast cancers resistant to anti-estrogen therapy, which correlates with elevated expression of NOTCH target genes HES1, HEY1 and HEY2, and is associated with increased population of breast cancer stem cells and distant metastases (Simoes et al. 2015). Development of int-3-induced mammary tumours in mice depends on Kit and Pdgfra signaling (Raafat et al. 2006) and on int-3-induced activation of NFkB signaling (Raafat et al. 2017). In head and neck squamous cell carcinoma (HNSCC), high NOTCH4 expression correlates with elevated HEY1 levels, increased cell proliferation and survival, epithelial-to-mesenchymal transition (EMT) phenotype and cisplatin resistance (Fukusumi et al. 2018). In melanoma, however, exogenous NOTCH4 expression correlates with mesenchymal-to-epithelial-like transition and reduced invasiveness (Bonyadi Rad et al. 2016). NOTCH4 is frequently overexpressed in gastric cancer. Increased NOTCH4 levels correlate with activation of WNT signaling and gastric cancer progression (Qian et al. 2015).

NOTCH4 is expressed in adipocytes and may promote adipocyte differentiation (Lai et al. 2013).

During Dengue virus infection, DLL1, DLL4, NOTCH4 and HES1 are upregulated in interferon-beta (INFB) dependent manner (Li et al. 2015). NOTCH4 signaling may be affected by Epstein-Barr virus (EBV) infection, as the EBV protein BARF0 binds to NOTCH4 (Kusano and Raab-Traub 2001).

Literature references

- Gutkind, JS., Sakai, A., Liu, C., Guo, TW., Califano, JA., Fukusumi, T. et al. (2018). The NOTCH4-HEY1 Pathway Induces Epithelial-Mesenchymal Transition in Head and Neck Squamous Cell Carcinoma. *Clin. Cancer Res.*, 24, 619-633. [↗](#)
- Rossant, J., Kitajewski, J., Uyttendaele, H., Ho, J. (2001). Vascular patterning defects associated with expression of activated Notch4 in embryonic endothelium. *Proc. Natl. Acad. Sci. U.S.A.*, 98, 5643-8. [↗](#)
- Bargo, S., Callahan, R., Raafat, A., Anver, MR. (2004). Mammary development and tumorigenesis in mice expressing a truncated human Notch4/Int3 intracellular domain (h-Int3sh). *Oncogene*, 23, 9401-7. [↗](#)
- Kusano, S., Raab-Traub, N. (2001). An Epstein-Barr virus protein interacts with Notch. *J. Virol.*, 75, 384-95. [↗](#)
- MacKenzie, F., Wong, F., Karsan, A., Duriez, P., Nosedá, M. (2004). Notch4 inhibits endothelial apoptosis via RBP-Jkappa-dependent and -independent pathways. *J. Biol. Chem.*, 279, 11657-63. [↗](#)

Editions

2004-12-15	Authored	Jassal, B.
2004-12-15	Reviewed	Joutel, A.
2012-02-11	Edited	Orlic-Milacic, M.
2018-04-05	Authored, Revised	Orlic-Milacic, M.
2018-05-01	Reviewed	Haw, R.
2018-05-09	Edited	Orlic-Milacic, M.

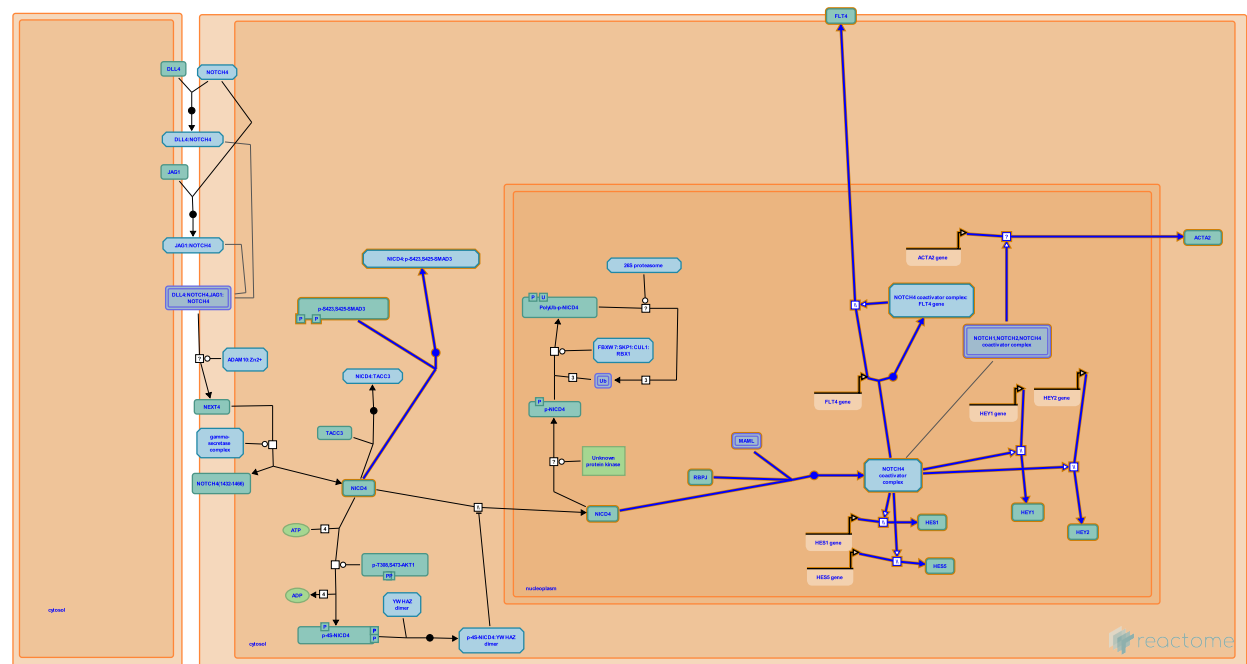
Editions

2018-04-05	Authored	Orlic-Milacic, M.
2018-05-01	Reviewed	Haw, R.
2018-05-09	Edited	Orlic-Milacic, M.

NOTCH4 Intracellular Domain Regulates Transcription ↗

Location: Signaling by NOTCH4

Stable identifier: R-HSA-9013695



In the nucleus, NOTCH4 intracellular domain fragment (NICD4) binds transcription factors RBPJ (CSL) and mastermind family members (MAML1, MAML2 or MAML3) to form the NOTCH4 co-activator complex (Lin et al. 2002). The NOTCH4 coactivator complex stimulates transcription of well-established NOTCH targets HES1, HES5, HEY1 and HEY2 in a cellular context-dependent manner (Lin et al. 2002, Raafat et al. 2004, Tsunematsu et al. 2004, Bargo et al. 2010). NOTCH4 also stimulates transcription of the FLT4 (VEGFR3) gene, encoding vascular endothelial growth factor receptor-3 (Shawber et al. 2007) and ACTA2 gene, encoding smooth muscle alpha actin (Tang et al. 2008).

NICD4 inhibits TGF-beta-induced SMAD-mediated transcription via binding of NICD4 to TGF-beta activated SMAD3 (Sun et al. 2005, Grabias and Konstantopoulos 2013).

Literature references

Konstantopoulos, K., Grabias, BM. (2013). Notch4-dependent antagonism of canonical TGF- β 1 signaling defines unique temporal fluctuations of SMAD3 activity in sheared proximal tubular epithelial cells. *Am. J. Physiol. Renal Physiol.*, 305, F123-33. ↗

Tsunematsu, R., Oike, Y., Bessho, Y., Hatakeyama, S., Nakayama, KI., Suda, T. et al. (2004). Mouse Fbw7/Sel-10/Cdc4 is required for notch degradation during vascular development. *J. Biol. Chem.*, 279, 9417-23. ↗

Oyama, T., Nagase, T., Lin, SE., Kitagawa, M., Harigaya, K. (2002). Identification of new human mastermind proteins defines a family that consists of positive regulators for notch signaling. *J. Biol. Chem.*, 277, 50612-20. ↗

Liaw, L., Tang, Y., Urs, S. (2008). Hairy-related transcription factors inhibit Notch-induced smooth muscle alpha-actin expression by interfering with Notch intracellular domain/CBF-1 complex interaction with the CBF-1-binding site. *Circ. Res.*, 102, 661-8. ↗

Feirt, N., Vorontchikhina, M., Chawengsaksophak, K., Borisenko, V., Stowell, SA., Shiraishi, K. et al. (2007). Notch alters VEGF responsiveness in human and murine endothelial cells by direct regulation of VEGFR-3 expression. *J. Clin. Invest.*, 117, 3369-82. ↗

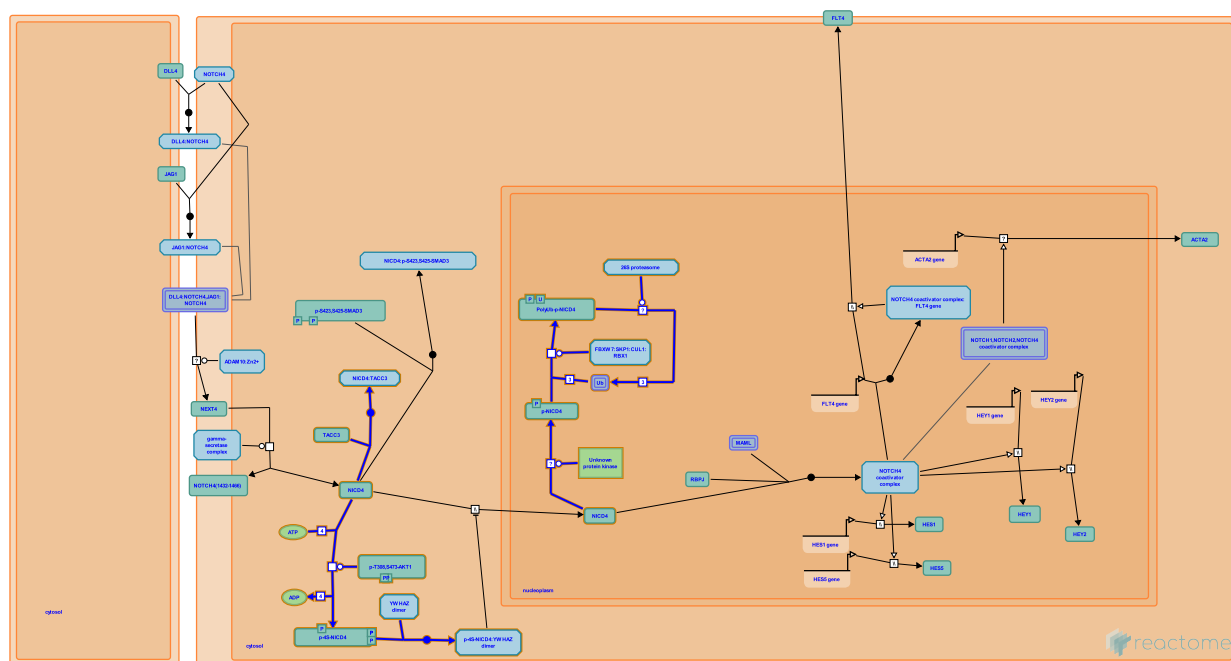
Editions

2018-04-05	Authored	Orlic-Milacic, M.
2018-05-01	Reviewed	Haw, R.
2018-05-09	Edited	Orlic-Milacic, M.

Negative regulation of NOTCH4 signaling ↗

Location: Signaling by NOTCH4

Stable identifier: R-HSA-9604323



NOTCH4 signaling can be negatively regulated at the level of nuclear translocation of the NOTCH4 intracellular domain fragment (NICD4). AKT-mediated phosphorylation of NICD4 promotes binding of NICD4 with 14-3-3-zeta (YWHAZ), leading to retention of NICD4 in the cytosol (Ramakrishnan et al. 2015).

The E3 ubiquitin ligase FBXW7, a component of the SCF ubiquitin ligase complex, binds to and ubiquitinates phosphorylated NICD4, targeting it for proteasome-mediated degradation (Wu et al. 2001). The level of NICD4 is significantly increased in Fbxw7 knockout mouse embryos, which die in utero and have impaired development of the vascular system (Tsunematsu et al. 2004).

Binding of NOTCH4 to ELOC (elongin C) is involved in proteasome-mediated degradation of NOTCH4, but the exact mechanism has not been elucidated (Cummins et al. 2011). MDM2, a TP53-induced ubiquitin ligase, was reported to ubiquitinate NICD4 and target it for degradation in response to TP53 activation (Sun et al. 2011).

NOTCH4 signaling is inhibited by binding of NICD4 to the transforming acidic coiled-coil protein-3, but the mechanism is not known (Bargo et al. 2010).

Literature references

- Tsunematsu, R., Oike, Y., Bessho, Y., Hatakeyama, S., Nakayama, KI., Suda, T. et al. (2004). Mouse Fbw7/Sel-10/Cdc4 is required for notch degradation during vascular development. *J. Biol. Chem.*, 279, 9417-23. ↗
- Green, AR., Davaakhuu, G., Pannuti, A., Ramakrishnan, G., Zhu, H., Chung, WC. et al. (2015). AKT and 14-3-3 regulate Notch4 nuclear localization. *Sci Rep*, 5, 8782. ↗
- Cummins, TD., Korte, EA., Khundmiri, SJ., Klein, JB., Barati, MT., Powell, DW. et al. (2011). Elongin C is a mediator of Notch4 activity in human renal tubule cells. *Biochim. Biophys. Acta*, 1814, 1748-57. ↗
- Das, I., Wu, G., Deshaies, RJ., Kitajewski, J., Li, J., Pauley, A. et al. (2001). SEL-10 is an inhibitor of notch signaling that targets notch for ubiquitin-mediated protein degradation. *Mol Cell Biol*, 21, 7403-15. ↗
- Lee, JM., Klauzinska, M., Sun, Y., Callahan, R., Santopietro, S., Artavanis-Tsakonas, S. et al. (2011). Trp53 regulates Notch 4 signaling through Mdm2. *J. Cell. Sci.*, 124, 1067-76. ↗

Editions

2018-04-05	Authored	Orlic-Milacic, M.
2018-05-01	Reviewed	Haw, R.
2018-05-09	Edited	Orlic-Milacic, M.

Table of Contents

Introduction	1
❖ Signaling by NOTCH4	2
❖ NOTCH4 Activation and Transmission of Signal to the Nucleus	4
❖ NOTCH4 Intracellular Domain Regulates Transcription	6
❖ Negative regulation of NOTCH4 signaling	7
Table of Contents	9